

# BioComputing's Network-Graph Tool

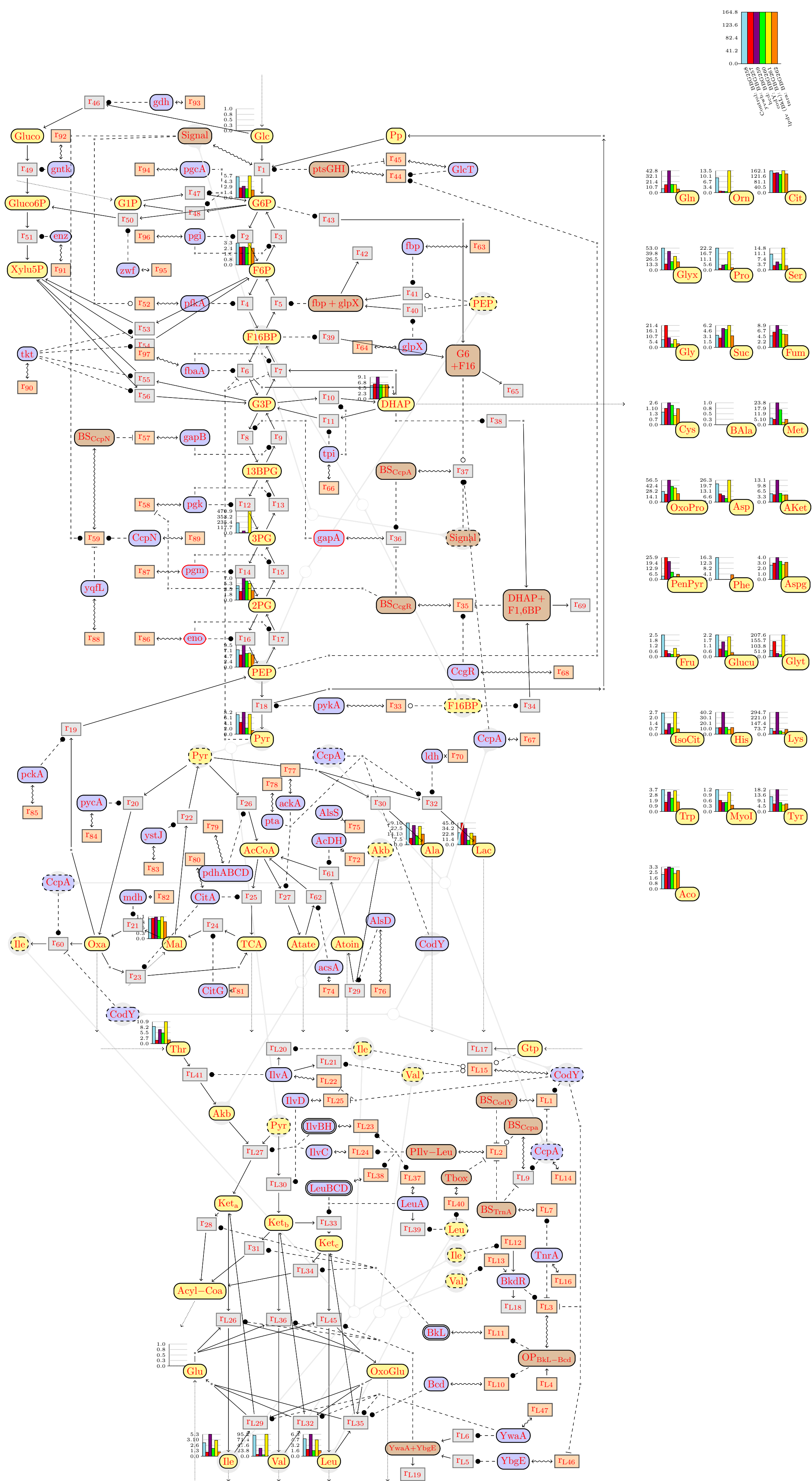
## Version 0.99

Joachim Niehren  
Inria & BioComputing, Lille, France

### 1 Reaction Network `Networks/b-subtilis-test.xml`

*Reaction Network.* See file: `Networks/b-subtilis-test.xml` See Figure 1.

*Analysis* `means-native-24h`



**Fig. 1.** The Networks/b-subtilis-test.xml.

Name	Function
r1	Phosphorylation of G
r1'	degradation of Signal
r2	Catalyse the conversion of G6P to F6P
r3	Catalyse the conversion of F6P to G6P ?
r4	Catalyse the conversion of F6P to F16BP
r5	Catalyse the conversion of F16BP to F6P
r6	Conversion of F16BP to G3P
r7	Conversion of G3P to F16BP
r8	Catalyse the conversion of G3P to 13BPG
r9	Catalyse the conversion of 13BPG to G3P
r10	Catalyse the conversion of G3P to DHAP
r11	Catalyse the conversion of DHAP to G3P
r12	Catalyse the conversion of 13BPG to 3PG
r13	Catalyse the conversion of 3PG to 13BPG
r14	Catalyse the conversion of 3PG to 2PG
r15	Catalyse the conversion of 2PG to 3PG
r16	Catalyse the conversion of 2PG to PEP
r17	Catalyse the conversion of PEP to 2PG
r18	Catalyse the conversion of PEP to Pyr
r33	expression of pykA
r33'	degradation of pykA
r34	Express F16BP
r35	Bind CcgR to gapA for inhibition
r35'	degradation of BS <sub>CcgR</sub>
r36	Regulation of gapA activity, activation by CcpA and inhibition by CcgR
r36'	degradation of gapA
r37	Bind CcpA to gapA for activation
r37'	degradation of BS <sub>CcpA</sub>
r38	Express DHAP
r39	Expression of F16BP
r40	Express glpX
r41	Express fbp
r42	degradation of fbp + glpX
r43	Express G6P
r44	Activation of GlcT activity
r44'	degradation of ptsGHI
r45	Inhibition of GlcT activity by pstGHI
r45'	degradation of GlcT
r46	Production of G throw Gluco
r47	Catalyse the conversion of G1P to G6P
r48	Catalyse the conversion of G6P to G1P
r49	Production of Gluco throw Gluco6P
r50	Production of G6P throw Gluco6P
r51	Production of Xylu5P throw Gluco6P
r52	expression of pfkA
r52'	degradation of pfkA
r53	Production of Xylu5P throw F6P
r54	Production of Xylu5P throw F6P
r55	Production of G3P throw Xylu5P
r56	Production of Xylu5P throw G3P
r57	Expression of gapB
r57'	degradation of gapB
r58	expression of pgk
r58'	degradation of pgk
r59	Bind CcpN to gapB for inhibition
r59'	degradation of BS <sub>CcpN</sub>
r63	expression of fbp
r63'	degradation of fbp
r64	expression of glpX
r64'	degradation of glpX
r65	degradation of G6 +F16
r66	degradation of tpi
r66'	degradation of tpi
r67	expression of CcpA
r67'	degradation of CcpA
r68	expression of CcgR
r68'	degradation of CcgR
r69	degradation of G6P + DH + F2
r86	expression of eno
r86'	degradation of eno
r87	expression of pgm
r87'	degradation of pgm
r88	degradation of yqfL
r88'	degradation of yqfL
r89	expression of CcpN
r89'	degradation of CcpN
r90	degradation of tkt
r90'	degradation of tkt
r91	degradation of enz
r91'	degradation of enz
r92	degradation of gntk
r92'	degradation of gntk
r93	degradation of gdh
r93'	degradation of gdh
r94	expression of pgcA
r94'	degradation of pgcA
r95	degradation of zwf
r95'	degradation of zwf
r96	expression of pgi
r96'	degradation of pgi

r97	expression of fbaA
r97'	degradation of fbaA
r20	Production of Atoin throw Pyr
r21	Production of Oxa throw Mal
r22	Production of Pyr throw Mal
r23	Production of TCA throw Oxa
r24	Production of Mal throw TCA
r25	Turn to TCA throw AcCoA
r26	Production of AcCoA from Pyr
r27	Production of Atate from AcCoA
r29	Production of Atoin from Aceto
r30	Production of Alanine from Pyr
r32	Production of Lac from Pyr
r60	Production of Ile from Oxa
r61	Production of AcCoA from Atoin
r62	Production of AcCoA from Atate
r70	expression of ldh
r70'	degradation of ldh
r72	expression of AcDH
r72'	degradation of AcDH
r74	expression of acsA
r74'	degradation of acsA
r75	expression of AlsS
r75'	degradation of AlsS
r76	expression of eno
r76'	degradation of AlsD
r77	expression of ackA
r77'	degradation of ackA
r78	expression of pta
r78'	degradation of pta
r79	expression of pdhABCD
r79'	degradation of pdhABCD
r80	expression of CitA
r80'	degradation of CitA
r81	expression of CitG
r81'	degradation of CitG
r82	expression of mdh
r82'	degradation of mdh
r83	expression of ystJ
r83'	degradation of ystJ
r84	expression of pycA
r84'	degradation of pycA
r85	expression of pckA
r85'	degradation of pckA
r19	feedback reaction to PEP from Oxa
rL1	bind CodY to Pilv–Leu for inhibition
	predition: single knockout, unsafe
	experiment: OK (we have this mutant)
rL1'	degradation of BS <sub>CodY</sub>
rL2	activate Pilv–Leu promoter
	predition: no
	experiment: not done, because we need the expression of Pilv – Leu
	in order to activate the reaction r23, r24, r39, r37
rL2'	degradation of Pilv–Leu
rL3	bind BkdR to BkL Bcd promoter
	experiment: Ok
rL3'	degradation of OP <sub>BkL–Bcd</sub>
rL4	constitutive expression of BkL Bcd operon
	experiment: Ok
rL5	express YbgE
rL6	express YwaA
rL7	bind TnrA to Pilv–Leu promoter for inhibition
	predition: safe single knockout
	experiment: OK (we have this mutant)
rL7'	degradation of BS <sub>TnrA</sub>
rL9	bind CcpA to Pilv–Leu promoter without BS <sub>TnrA</sub> loop
rL9'	degradation of BS <sub>CcpA</sub>
rL10	expression of Bcd, activated by OP <sub>BkL–Bcd</sub>
	experiment: OK (we have this mutant)
rL10'	degradation of Bcd
rL11	expression of BkL, activated by OP <sub>BkL–Bcd</sub>
	experiment: OK (we have this mutant)
rL11'	degradation of BkL
rL12	activate BkdR by Ile
	experiment: Francois: I need to check what is the mechanism of this activation.
	If is via the Val-tRNA is not possible because the gene
	encoded for the valyl-tRNA synthetase is essential in B. subtilis
rL13	activate BkdR by Val
	experiment: Francois: I need to check what is the mechanism of this activation
	if it is via the Ile-tRNA is not possible because the gene encoded
	for the isoleucyl-tRNA synthetase is essential in B. subtilis
rL14	expression of CcpA
	predition: single-knockout, unsafe
	experiment: It seems to be possible but we do not see the interest since CcpA is
	an activator of Pilv – Leu and a repressor of CodY
rL14'	degradation of CcpA
rL15	express and accelerate CodY (to be explained acceleration)
	OK (we have this mutant)
rL15'	degradation of CodY
rL16	expression of TnrA
	experiment: OK (we have this mutant)
rL16'	degradation of TnrA
rL17	Gtp degradation
rL18	BkdR degradation
rL19	YwaA+YbgE degradation
rL20	disabeling of IlvA by Ile
rL21	disabeling of IlvA by Val
rL22	expression of IlvA inhibited by binding of CodY

	experiment: Ok but the <b>IlvA</b> expression is <b>r21</b> ? (we have this mutant but the result were not good)
<b>rL22'</b>	degradation of <b>IlvA</b>
<b>rL23</b>	expression of <b>IlvBH</b>
	experiment: NO, because we need the expression of <b>IlvBH</b> in order to activate the reaction <b>r30</b> , thereby producing <b>Ketb</b> , the precursor of <b>Ket<sub>c</sub></b>
<b>rL23'</b>	degradation of <b>IlvBH</b>
<b>rL24</b>	expression of <b>IlvC</b>
	experiment: NO, because we need the expression of <b>IlvC</b> in order to activate the reaction <b>r30</b> , thereby producing KetB the precursor of <b>Ket<sub>c</sub></b>
<b>rL24'</b>	degradation of <b>IlvC</b>
<b>rL25</b>	expression of <b>IlvD</b> , inhibited by binding of <b>CodY</b> to promotor
	experiment: NO, because we need the expression of <b>IlvD</b> in order to activate the reaction <b>r30</b> , thereby producing <b>Ketb</b> , the precursor of <b>Ket<sub>c</sub></b>
<b>rL25'</b>	degradation of <b>IlvD</b>
<b>rL26</b>	metabolic transformation of <b>Keta</b> to <b>Ile</b> activated by <b>YwaA+YbgE</b> , after an amino addition taken from <b>Glu</b> , which becomes <b>OxoGlu</b> .
<b>rL27</b>	metabolic transformation of <b>Akb</b> and <b>Pyr</b> into <b>Keta</b>
<b>r28</b>	prepare output of <b>Keta</b> activated by <b>BkL</b>
<b>rL29</b>	degradation of <b>Ile</b> into <b>Keta</b> activated by <b>YwaA</b> and <b>Bcd</b> , with an amino transfer from <b>OxoGlu</b> which becomes <b>Glu</b> .
<b>rL30</b>	metabolic transformation from <b>Pyr</b> to <b>Ketb</b> , activated by <b>IlvD</b>
<b>r31</b>	prepare output of <b>Ketb</b> activated by <b>BkL</b>
<b>rL32</b>	degradation of <b>Val</b> into <b>Ketb</b> activated by <b>YwaA</b> and <b>Bcd</b> with an amino transfer from <b>OxoGlu</b> which becomes <b>Glu</b> .
<b>rL33</b>	metabolic transformation of <b>Ketb</b> into <b>Ket<sub>c</sub></b> , activated by <b>LeuBCD</b> and <b>LeuA</b>
<b>rL34</b>	prepare output of <b>Ket<sub>c</sub></b> activated by <b>BkL</b>
<b>rL35</b>	degradation of <b>Leu</b> into <b>Ket<sub>c</sub></b> ctivated by <b>YwaA</b> and <b>Bcd</b> with an amino transfer from <b>OxoGlu</b> which becomes <b>Glu</b> .
<b>rL36</b>	metabolic transformation of <b>Ketb</b> to <b>Val</b> activated by <b>YwaA+YbgE</b> , after an amino addition from <b>Glu</b> which becomes <b>OxoGlu</b>
<b>rL37</b>	expression of <b>LeuA</b>
	experiment: NO, because we need the expression of leuA in order to activate the reaction <b>r33</b> , thereby producing <b>Ket<sub>c</sub></b> the precursor of <b>Leu</b>
<b>rL37'</b>	degradation of <b>LeuA</b>
<b>rL38</b>	expression of <b>LeuBCD</b>
	experiment: NO, because we need the expression of leuBCD in order to activate the reaction <b>r33</b> , thereby producing <b>Ket<sub>c</sub></b> the precursor of <b>Leu</b>
<b>rL38'</b>	degradation of <b>LeuBCD</b>
<b>rL39</b>	deactivation of <b>LeuA</b> by <b>Leu</b>
<b>rL40</b>	Leucine attenuation
	experiment: OK (we have this mutant)
<b>rL40'</b>	degradation of <b>Tbox</b>
<b>rL41</b>	metabolic transformation of <b>Thr</b> into <b>Akb</b> using <b>IlvA</b>
<b>rL45</b>	metabolic transformation of <b>Ket<sub>c</sub></b> to <b>Leu</b> activated by <b>YwaA+YbgE</b> after an amino addition from <b>Glu</b> , which becomes <b>OxoGlu</b>
<b>rL46</b>	expression of <b>YbgE</b> , inhibited by binding of <b>CodY</b> to promoter
	experiment: NO, because we need the expression of <b>YbgE</b> in order to activate the reaction <b>r5</b> , which produce <b>Leu</b>
<b>rL46'</b>	degradation of <b>YbgE</b>
<b>rL47</b>	expression of <b>YwaA</b>
<b>rL47'</b>	experiment: Ok degradation of <b>YwaA</b>

**Fig. 3.** Reactions of Networks/b-subtilis-test.xml

### 1.1 What Else

*Comments to be treated* A small FAQ

Question 1. Are r40 r41 candidates for KO ?

Question 2. Define Signal real name

Question 3. Do we add G1P outflow ? do we remove the inflow ? is pgcA an Accelerator ?

Question 4. Precision for CcpA from Tobish 1999?

Question 5. Repression of CodY by Val-Leu (r71)? (cf Carbon Catabolic Control of the Metabolic Network in B. Subtilis)

Question 6. Repression of production of Malate throw TCA by CcpA (r24)? (cf Positive regulation of B. Subtilis by CodY ... )

Question 7. Ilv actor meaning ?

A small FAQ

A small FAQ

Question 2. Why is reaction **r3** needed, given that **r4** produces **OP<sub>BkL-Bcd</sub>** anyway? Answer: since more **OP<sub>BkL-Bcd</sub>** is produced this way.

Question 3. Is **Akb** input from context needed, since **Akb** can also be produced from **Thr** (**r9**)? Answer: yes since external pathways can produce it too.



Role	Short name	Chemical Species
Metabolites	<b>Glc</b>	D-Glucose
	<b>G1P</b>	Glucose-1-Phosphate
	<b>G6P</b>	Glucose-6-Phosphate
	<b>F6P</b>	Fructose-6-Phosphate
	<b>F16BP</b>	Fructose-1,6-Biphosphate
	<b>G3P</b>	Glyceraldehyde-3-Phosphate
	<b>13BPG</b>	1,3-Bisphosphoglycerate
	<b>3PG</b>	3-Phosphoglycerate
	<b>2PG</b>	2-Phosphoglycerate
	<b>PEP</b>	Phosphoenolpyruvate
	<b>Pyr</b>	Pyruvate
	<b>DHAP</b>	Dihydroxyacetonphosphate
	<b>Pp</b>	PyroPhosphate
	<b>Gluco</b>	Gluconate
	<b>Gluc6P</b>	Glucose-6-P
	<b>Xylu5P</b>	Xylulose-5-P
	<b>TCA</b>	Krebs cycle
	<b>Atoin</b>	Acetoin
	<b>Atate</b>	Acetate
	<b>Ala</b>	Alanine
	<b>Lac</b>	Lactate
	<b>AcCoA</b>	Acetyl CoA
	<b>Oxa</b>	Oxaloacetate
	<b>Mal</b>	D-Malate
	<b>Ile</b>	Isoleucine
	<b>Leu</b>	Leucine
	<b>Val</b>	Valine
	<b>Akb</b>	L-2-amino-acetoacetate
	<b>Glu</b>	L-Glutamate
	<b>OxoGlu</b>	Oxogluterate
	<b>Gtp</b>	Guanosine triphosphate
	<b>Ket<sub>a</sub></b>	2-keto-3-methylvalerate
	<b>Acyl-Coa</b>	Acyl Coenzyme A
	<b>Ket<sub>b</sub></b>	2-keto-isovalerate
	<b>Ket<sub>c</sub></b>	2-keto-isocaproate
	<b>Thr</b>	Threonine deshydratase
	<b>Gln</b>	L-Glutamine
	<b>Orn</b>	L-Ornithine
	<b>Cit</b>	Citrate
	<b>Glyx</b>	Glyoxalat
	<b>Pro</b>	L-Proline
	<b>Ser</b>	L-Serine
	<b>Gly</b>	Glycine
	<b>Suc</b>	Succinate
	<b>Fum</b>	Fumarate
	<b>Cys</b>	L-Cysteine
	<b>BAla</b>	Beta-alanine
	<b>Met</b>	L-Methionine
	<b>OxoPro</b>	5-oxoproline
	<b>Asp</b>	Aspartate
	<b>AKet</b>	Alpha ketoglutaric acid
	<b>PenPyr</b>	Penylpyruvate
	<b>Phe</b>	L-Phenylalanine
	<b>Aspg</b>	L-Asparagine
	<b>Fru</b>	Fructose
	<b>Glucu</b>	Glucuronate
	<b>Glyt</b>	Glycerate
	<b>IsoCit</b>	Isocitrate
	<b>His</b>	L-Histidine
	<b>Lys</b>	L-Lysine
	<b>Trp</b>	L-Tryptophane
	<b>MyoI</b>	Myo-inositol
	<b>Tyr</b>	Tyrosine
	<b>Aco</b>	Trans-aconicic acid
Proteines	<b>GlcT</b>	Transcriptional antiterminator.
	<b>pgi</b>	Glu-6-Phosphate isomerase.
	<b>pfkA</b>	Phosphofructokinase.
	<b>fbaA</b>	Fructose-1,6-biphosphate aldolase
	<b>gapB</b>	Glyceraldehyde-phosphatedehydrogenase
	<b>pgk</b>	Phosphoglycerate kinase
	<b>pgm</b>	2,3-Biphosphoglycerate - Independent phosphoglycerate mutase
	<b>eno</b>	Enolase
	<b>pykA</b>	Pyruvate kinase
	<b>tpi</b>	Triose phosphate isomerase
	<b>gapA</b>	Glyceraldehyde-3-phosphate dehydrogenase
	<b>CcgR</b>	Transcriptional repressor
	<b>CcpN</b>	Transcriptional repressor
	<b>yqfL</b>	Positive regulator
	<b>fbp</b>	Fructose-1,6-biphosphatase class III
	<b>glpX</b>	Fructose-1,6-biphosphatase class II
	<b>pgcA</b>	$\alpha$ -phosphoglucomutase
	<b>gdh</b>	Glucose 1-deshydrogenase
	<b>gntk</b>	Gluconokinase
	<b>enz</b>	??
	<b>tkt</b>	Transketolase
	<b>zwf</b>	Glucose-6-phosphate 1-dehydrogenase
	<b>CcpA</b>	Transcriptional activator
	<b>pdhABCD</b>	Pyruvate dÃfshydrogenase
	<b>ystJ</b>	Enzyme malique
	<b>CitA</b>	Citrate synthase
	<b>CitG</b>	Fumarase
	<b>mdh</b>	Malate deshydrogenase
	<b>pckA</b>	Phosphoenolpyruvate carboxykinase
	<b>pycA</b>	Pyruvate carboxylase
	<b>ackA</b>	Acetate kinase
	<b>pta</b>	Phosphotransacetylase
	<b>AcDH</b>	Acetoin deshydrogenase
	<b>acsA</b>	Acetyl CoA synthetase
	<b>AlsS</b>	$\alpha$ -acetolactate synthetase
	<b>AlsD</b>	$\alpha$ -acetolactate-dehydrogenase
	<b>ldh</b>	Lactate dehydrogenase
	<b>CodY</b>	Transcriptional pleiotropic regulator
	<b>Bcd</b>	Branched chain amino-acid dehydrogenase
	<b>BkL</b>	2-oxoisovalerate dehydrogenase
	<b>BkdR</b>	Transcriptional activator of <b>BkL</b>
	<b>IlvA</b>	Threonine deshydratase
	<b>IlvBH</b>	Acetolactate synthase
	<b>IlvC</b>	Ketol-acid reductoisomerase
	<b>IlvD</b>	Dihydroxy-acid dehydratase
	<b>LeuA</b>	2-isopropylmalate synthase
	<b>LeuBCD</b>	3-isopropylmalate dehydratase
	<b>TnrA</b>	Nitrogen pleiotropic transcrptional regulator
	<b>YbgE</b>	Branched chain amino-acid aminotransferase
	<b>YwaA</b>	branched chain amino-acid aminotransferase
Actors	<b>ptsGHI</b>	Composed by ptsG, ptsH and ptsI
	<b>fbp + glpX</b>	Activity of fbp and glpX
	<b>DHAP+ F1,6BP</b>	Activity of G6P, DHAP, F6P and F16BP
	<b>G6 +F16</b>	Activity of <b>G6P</b> and <b>FOneSixBP</b>
	<b>BS<sub>CcpA</sub></b>	Activity of <b>CcpA</b> binding to <b>gapA</b>
	<b>BS<sub>CcgR</sub></b>	Activity of <b>CcgR</b> binding to <b>gapA</b>
	<b>BS<sub>CcpN</sub></b>	Activity of <b>CcpN</b> binding to <b>gapB</b>
	<b>Signal</b>	Signal generated by the phosphorylation of <b>G</b> to <b>G6P</b>
	<b>Pilv-Leu</b>	Activity of promoter of <b>IlvBH IlvC LeuA LeuBCD</b> operon
	<b>BS<sub>CodY</sub></b>	Activity of <b>CodY</b> binding to promoter <b>PilvLeu</b>
	<b>BS<sub>TnrA</sub></b>	Activity of <b>TnrA</b> binding to promoter <b>PilvLeu</b>
	<b>BS<sub>CcpA</sub></b>	Activity of <b>CcpA</b> binding to promoter <b>PilvLeu</b> without <b>BS<sub>TnrA</sub></b> loop
	<b>OP<sub>BkL-Bcd</sub></b>	Activity of promoter of <b>BkL Bcd</b> operon
	<b>YwaA+YbgE</b>	Activity of <b>YbgE</b> and <b>YwaA</b>
	<b>Tbox</b>	activity of tryptophan attenuation

**Fig. 2.** Molecules of Networks/b-subtilis-test.xml.