

QuantPred

Quantitative Change Prediction for Reaction Networks

PhD Proposal 2022

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Abstract

We propose to develop novel bioinformatics methods in area of systems biology to improve the production of microbial biosurfactants with methods from biotechnology. Biosurfactants produced by microorganisms have the potential to replace chemical pesticides in agriculture, to serve for detergency industry and for cleaning petrol on the sea. These applications are relevant for the planet, so it is of highest interest to enable the production of biosurfactants of better quality, in higher quantities, and at lower prices.

Nonribosomal lipopeptides are a class of microbial biosurfactants that can be produced from amino and fatty acids by many bacteria such as *Bacillus* and *Pseudomonas*. In our previous interdisciplinary work we have developed successful methods for overproducing surfactine in the bioreactor based on artificial strains of *Bacillus subtilis* obtained by out-knocking dedicated genes. While the case of single gene knockouts has been largely studied, we believe that multiple gene knockouts are the key for better optimization methods in the future. Multiple gene knockout, however, raise serious difficulties for exhaustive tests in the wet lab, simply since the possibly number is too large.

The hope of model-based prediction methods from systems biology is to limit the number of candidates, so that only few of the many possible candidates need to be tested experimentally. The restriction to purely qualitative reasoning techniques, however, makes it difficult if not impossible to distinguish the good multiple knockouts from the others. Therefore, we propose to add quantitative aspects to existing prediction methods and to the existing biochemical reaction networks of the metabolism of *B. subtilis* and its control. Of course, this is not generally possible, since only much of the quantitative information on the regulatory control is unknown and will remain unknown in the near future. We therefore propose to integrate known aspects on the forms of the kinetic functions (mass-action, Michaelis-Menten, different kinds of repression, etc), both into the formal models of the metabolic and control networks and into model-based prediction methods.

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A. Organisation

A.1. Interdisciplinary Research Teams

BioComputing team Institut Charles Violette	Université de Lille Université de Lille	Cristal lab INRAE UMR transfrontalière BioEcoAgro (1158)
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The INRAE UMR transfrontalière BioEcoAgro headed by Philippe JACQUES, subsumes several labs of the Haut de France, including the Laboratory Charles Violette of the University of Lille, and the TERRA lab of the Université de Liège in Gembloux.

- the bioinformatics group BioComputing works systems biology
- our partner Francois Coutte from BioEcoAgro works on biotechnology.

A.2. Funding Plans

Required 50%, Université de Lille

Co-funding 50% ANR Migad possible

A.3. Related Funded Projects

ANR Migad the topic subscribes to aspects the ANR Project Migad (2021-2025) coordinated by Cédric Lhoussaine (BioComputing). Cristian Versari and Joachim Niehren are leading the research on these aspects of Migad.

AI the research proposal promised to develop logical methods from classical artificial intelligence for representing the knowledge of biological systems and reasoning with it.

A.4. Priorities of Region Haut de France

CPER BiHautsEco the biotechnology aspect of QuantPred subscribes to the regions CPER on biotechnology for agro-economics. This CPER is coordinated by the co-director of the proposed PhD thesis, Philippe JACQUES.

CPER CornellIA the computer science aspect of QuantPred subscribes to the regions CPER on artificial intelligence.

Both CPERs are supported by the **ISite** of the Université de Lille.

B. History of the Cooperation

This interdisciplinary cooperation was started by Joachim Niehren and F. Coutte via the postdoc of Mathias John in the BioComputing group (2011-2013) see [2,3,4,6,7,8]. The cooperation was continued with on the biotechnology side the PhD theses of Debarun Dhali (2014-2017) in cooperation with F. Coutte see [5], the PhD project of Ariane Theatre (2019-2021) see [9], and on the bioComputing side by the PhD thesis and postdoc of Emily Allart (2016-2021) [-1,0,1]. In 2022, it led publication at CMSB, the international conference on systems biology [-3] and to an interdisciplinary publication at the biotechnology journal *Metabolites* [-2].

The cooperation was supported through ERA CoBioTech, H2020 project BestBioSurf (2018-2022). Parts of the project subscribe to the ANR project Migad that is coordinated by the head of BioComputing's C. Lhoussaine.

C. State of the Art

Microbial biosurfactants have the potential to replace chemical pesticides in agriculture, chemical cleaners part in detergency industry, and to serve for cleaning petrol on the sea. All three applications are relevant for the planet, so it is of highest interest to enable the production of biosurfactants of better quality, in higher quantities, and at lower prices. Surfactin is a nonribosomal lipopeptide that can be produced from amino and fatty acids by many bacteria such as *Bacillus* and *Pseudomonas* [14, 15]. Surfactin is one of the most powerful biosurfactants known and it displays several biological activities of interest (antiviral, antimycoplasmic, elicitor, etc). It consists of a ring that carries 7 amino acids of which 4 are leucine and one fatty acid chain of different length and isomers. It is assembled by a non ribosomal mechanism involving multifunctional proteins called Non Ribosomal Peptide Synthesis (NRPS). The overproduction of surfactin on [13] was obtained by replacing the native promoter of the surfactin operon (*srfA*) by a constitutive one. This led to new artificial strains of *B. Subtilis*, which then are overproducing surfactin in the bioreactor.

Leucine is a branched chain amino acid produced by a metabolic pathway with complex regulatory mechanisms. The case of knockouts of single genes that are relevant for the production of leucine has been studied in our previous work. For this, we formally modelled the reaction network of leucine production, and developed novel qualitative reasoning methods based on formal methods, that on abstract interpretation [1,0-4] and constraint programming. We also combined this approach with techniques from flux balance analysis [10, 11, 12]. This way we obtained 12 single knockout predictions for leucine overproduction of which 6 were confirmed by wet lab experiments [5-9]. Of course, these techniques for modelling and prediction are sufficiently general so that they should be useful for quite different applications. But still there also remains much space for improvements concerning the quantity, quality, and price of the surfactin production, that we would like to explore.

D. Research Project

Challenge. We believe that multiple gene knockouts are the key to better optimization methods in the future. Multiple gene knockout, however, are very difficult for exhaustive tests in the wet lab, simply since the possibly number too large. The hope of model-based prediction methods from systems biology is to limit the number of candidates, so that only few of the many possible candidates need to be tested experimentally. The restriction to purely qualitative reasoning techniques, however, makes it difficult if not impossible to distinguish the good multiple knockouts from the others.

Approach. Therefore, we propose to add quantitative aspects to existing prediction methods and to the existing biochemical reaction networks of the metabolism of *B. subtilis* and its control. Of course, this is not generally possible, since only much of the quantitative information on the regulatory control is unknown and will remain unknown in the near future. We therefore propose to integrate known aspects on the forms of the kinetic functions (mass-action, Michaelis-Menten, different kinds of transcriptomic induction or repression, etc), both into the formal models of the metabolic and control networks and into model-based prediction methods. Our approach to make quantitative predictions will be to use interval based methods instead of purely qualitative reasoning on increases and decreases. The intervals will abstract from size of changes. In particular, when it comes to Michaelis-Menten kinetics, one needs to know whether one is close to saturation or far away, in order to estimate the size of increase or decreases. How this kind of knowledge can be incorporated into the models and analysis is the topic of the present PhD project.

Working program

Year 1

In the first 6 months, the student has to learn the state of the art, methodologically and technologically. Therefore

A. we will revisit and improve the existing qualitative prediction methods based on qualitative reasoning, so that they become more precise. This will be tested on concrete challenges, where the models are already existing in the biocomputing group (leucine model, aspartate model). In the next 6 months we will work in parallel on modelling and analysis methods in parallel

B. We will study small parts of reaction network of leucine production, to see, which kind of quantitative information on the kinetics of the metabolic control could be added to the existing qualitative models, talking about activation and inhibition only

C. We will develop our ideas towards interval-based methods for qualitative prediction that can be applied to the example models from Year 1.B).

Year 2

We will continue with the 2 lines of the second half of the first year in parallel:

A. During a visit of the TERRA lab in Gembloux Belgium (with Philippe Jacques), the student will learn about biological modelling problems in the context of surfactin overproduction.

B. We will define a general modeling language that supports reaction networks with partial kinetic information and sufficient quantitative information to capture the example studied in year 1B).

C. We will develop and implement a general interval-based method for qualitative prediction of single knockouts and apply it to larger metabolic networks, that can be applied to all models of the language from Year 2.B).

Year 3

A. We will extend the prediction methods for single knockouts from Year 2c) to multiple gene knockouts and its application to a model of leucine and aspartate overproduction.

B. the student will write up his PhD thesis.

E. References

E.1. On previous work on bioinformatics and systems biology

- [3] **J. Niehren**, A. Vaginay, **C. Versari**: Abstract Simulation of Reaction Networks via Boolean Networks. CMSB: 20th International Conference on Computational Methods in Systems Biology. Springer. 2022
- [-2] J.S. Guez, **J. Niehren**, **F. Coutte**, P. Jacques et al.: Bioinformatics modelling and metabolic engineering of the branched chain amino acid pathway for specific production of mycosubtilin isoforms in *Bacillus subtilis*. *Metabolites*, 2022.
- [-1] E. Allart, **J. Niehren**, **C. Versari**: Computing Difference Abstractions of Linear Equation Systems. *Theoretical Computer Science*, Elsevier, volume 893: 72-104, 2021.
- [0] Allart E, **Niehren J**, **Versari C**. Exact Boolean Abstraction of Linear Equation Systems. *MDPI Computation*. 9(11):113, 2021.
- [1] E. Allart, **C. Versari**, **J. Niehren**: Computing Difference Abstractions of Metabolic Networks Under Kinetic Constraints. 17th International Conference on Computational Methods in Systems Biology (CMSB), Springer, 2019.
- [2] **J. Niehren**, **C. Versari**, M. John, F. Coutte, **P. Jacques**: Predicting Changes of Reaction Networks with Partial Kinetic Information. *BioSystems*, 2016.
- [3] **J. Niehren**, M. John, **C. Versari**, F. Coutte, **P. Jacques**: Qualitative Reasoning about Reaction Networks with Partial Kinetic Information. *Computational Methods for Systems Biology (CMSB)*, Springer, 2015
- [4] M. John, M. Nebut, **J. Niehren**: Knockout Prediction for Reaction Networks with Partial Kinetic Information. 14th International Conference on Verification, Model Checking, and Abstract Interpretation, Springer, 2013.

E.2. Own previous work on biochemistry and biotechnology

- [5] D. Dhali, F. Coutte, A. Argüelles Arias, S. Auger, V. Bidnenko, G. Chataigné, M. Lalk, **J. Niehren**, J. de Sousa, **C. Versari**, **P. Jacques**: Genetic engineering of the branched fatty acid metabolic pathway of *Bacillus subtilis* for the overproduction of surfactin C14 isoform. *Biotechnology Journal*, 2017.
- [6] F. Coutte, **J. Niehren**, D. Dhali, M. John, **C. Versari**, **P. Jacques**: Modeling Leucine's Metabolic Pathway and Knockout Prediction Improving the Production of Surfactin, a Biosurfactant from *Bacillus Subtilis*. *Biotechnology Journal*, 2015.
- [7] F. Coutte, M. John, M. Béchet, M. Nebut, **J. Niehren**, V. Leclère, **P. Jacques**: Synthetic Engineering of *Bacillus subtilis* to Overproduce Lipopeptide Biosurfactants. 9th European Symposium on Biochemical Engineering Science, 2012.
- [8] M. John, F. Coutte, M. Nebut, **P. Jacques**, **J. Niehren**: Knockout Prediction for Reaction Networks with Partial Kinetic Information: Application to Surfactin Overproduction in *Bacillus subtilis*. 3rd International Symposium on Antimicrobial Peptides, 2012.
- [9] A. Théâtre, C. Cano-Prieto, M. Bartolini, Y. Laurin, M. Deleu, **J. Niehren**, T. Fida, S. Gerbinet, M. Alanjary, M. Medema, A. Léonard, L. Lins, A. Arabolaza, H. Gramajo, H. Gross, **P. Jacques**: The surfactin-like lipopeptides from *Bacillus* spp.: natural biodiversity and synthetic biology for a broader application range. *Frontiers in Bioengineering and Biotechnology*, 2021.

E.3. External previous work on systems biology

- [10] E. Almaas, B. Kovacs, T. Vicsek, Z. N. Oltvai, and A. L. Barabasi. Global organization of metabolic fluxes in the bacterium *Escherichia coli*. *Nature*, 427:839–843, 2004.
- [11] Markus W Covert and Bernhard O Palsson. Constraints-based models: regulation of gene expression reduces the steady-state solution space. *J. Theor. Biol.*, 221(3):309–325, 2003.
- [12] Lope A Flórez, Katrin Gunka, Rafael Polanía, Stefan Tholen, and Jörg Stülke. Spabbats: A pathway-discovery method based on boolean satisfiability that facilitates the characterization of suppressor mutants. *BMC Syst. Biol.*, 5(1):5, 2011. Previous publications of our biotechnology partner
- [13] F Coutte, V Leclère, M Béchet, J-S Guez, D Lecouturier, M Chollet-Imbert, P Dhulster, and P Jacques. Effect of pps disruption and constitutive expression of srfa on surfactin productivity, spreading and antagonistic properties of *Bacillus subtilis* 168 derivatives. *J. Appl. Microbiol.*, 109(2):480–491, 2010.
- [14] P. Jacques. Surfactin and other lipopeptides from *Bacillus* spp. In *Biosurfactants*, pages 57–91. Springer, 2011.s for the efficient production of a lipopeptide pseudofactin by *Pseudomonas fluorescens* BD5. *Microbial cell factories*, 17(1), 121, 2018.
- [15] P Biniarz, F Coutte, F Gancel and M Łukaszewicz. High-throughput optimization of medium components and culture conditions.